

## Scott Carey

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**To:** Scott Carey  
**Subject:** NTRPA GB MEETING 11-03-2022 <><><> ITEM 2 <><><> PUBLIC COMMENT  
**Attachments:** Attorney Letter—Fisher.pdf; Tahoe Prosperity Center—Illegal Lobbying Letter.pdf; Limiting liability with positioning to minimize negative health effects of cellular phone towers.pdf; NATURE—Radiofrequency EMF irradiation effects on pre-B lymphocytes.pdf

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NTRPA Governing Board:

It is time to take a stand against overtourism. Every meeting should be seen as an opportunity for you to take on the poster child of overtourism: the placement of cell towers on or around our homes. The explanation for why the existing wireless facilities are never good enough always boils down to incessantly increasing tourist demand. We the residents of this town must not be collateral damage for that ignoble cause. Like most of my Tahoe friends, I live here to connect with nature, not to be violated by morally bankrupt entrepreneurial spirits. Avarice rampantly blinds some prominent business leaders (below), but don't be fooled yourself. Be on the right side of change, be on the right side of social and environmental justice, be on the right side of history.

Cell towers are the face of overtourism:

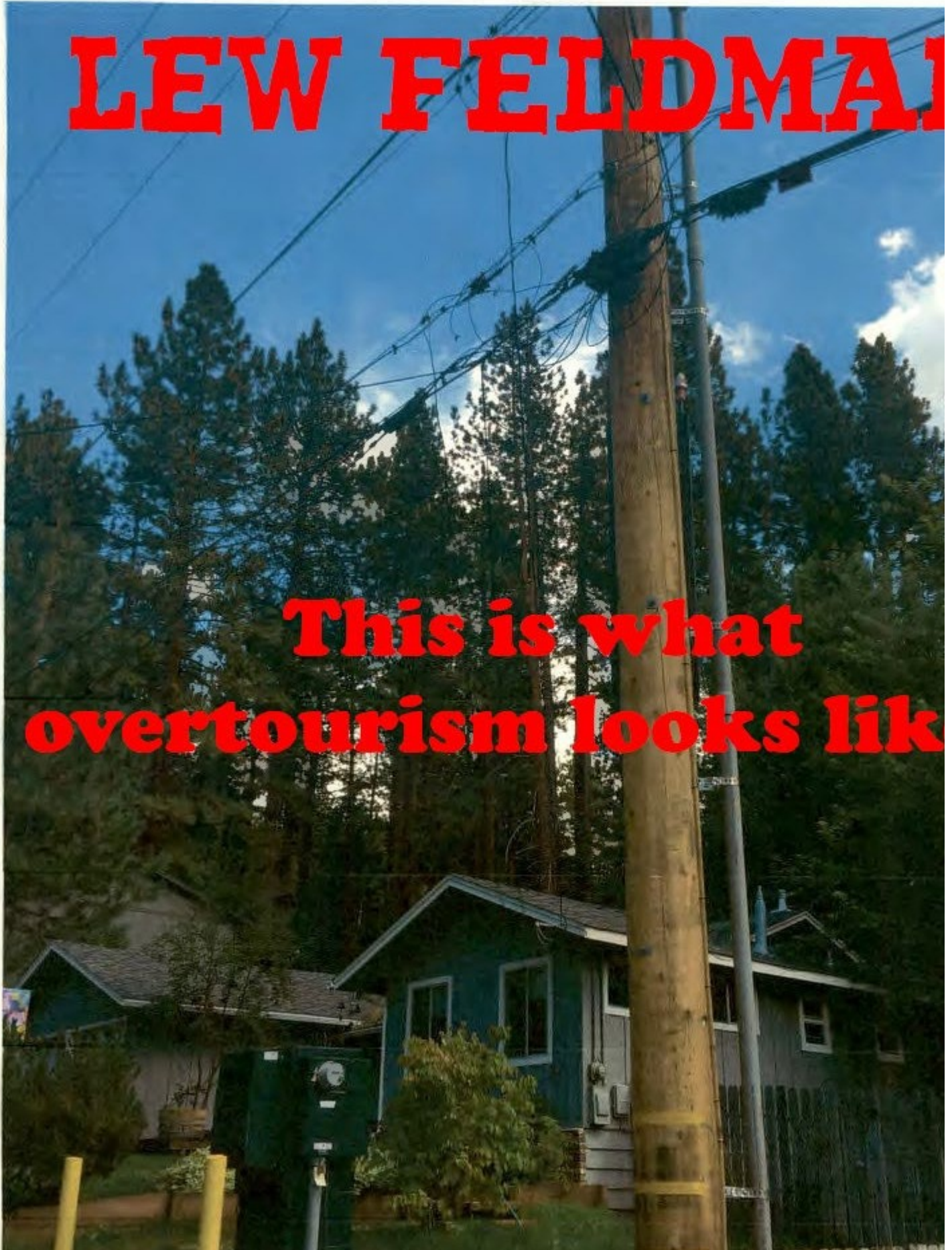
**Carol Chaplin**

**This is what  
overtourism**

and the boyfriend...

**LEW FELDMAN**

**This is what  
overtourism looks like**



Please see to it that you represent the community who elected you, not the out-of-city/state business interests that backed your campaign for office.

Best wishes,

Peter Norman







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# Limiting liability with positioning to minimize negative health effects of cellular phone towers

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## ABSTRACT

The use of cellular phones is now ubiquitous through most of the adult global population and is increasingly common among even young children in many countries (e.g. Finland, where the market for smart phones is nearly saturated). The basic operation of cellular phone networks demands widespread human exposure to radio-frequency radiation (RFR) with cellular phone base stations providing cellular coverage in most areas. As the data needs of the population increase from the major shift in the source of Internet use from personal computers to smart phones, this coverage is widely predicted to increase. Thus, both the density of base stations and their power output is expected to increase the global human RFR exposure. Although direct causation of negative human health effects from RFR from cellular phone base stations has not been finalized, there is already enough medical and scientific evidence to warrant long-term liability concerns for companies deploying cellular phone towers. In order to protect cell phone tower firms from the ramifications of the failed paths of other industries that have caused unintended human harm (e.g. tobacco) this Current Issue summarizes the peer-reviewed literature on the effects of RFR from cellular phone base stations. Specifically the impacts of siting base stations are closely examined and recommendations are made for companies that deploy them to minimize their potential future liability.

## 1. Negative human health effects from proximity to cellular phone base stations

There is a large and growing body of evidence that human exposure to RFR from cellular phone base stations causes negative health effects (Siddoo-Atwal, 2018; Singh et al., 2018; Faisal, et al., 2018) including both i) neuropsychiatric complaints such as headache, concentration difficulties, memory changes, dizziness, tremors, depressive symptoms, fatigue and sleep disturbance (Navarro et al., 2003; Hutter et al., 2006; Abdel-Rassoul et al., 2007); and ii) increased incidence of cancer and living in proximity to a cell-phone transmitter station (Wolf and Wolf, 2004; Havas, 2017). The mechanism for causing cancer could be from observed genetic damage using the single cell gel electrophoresis assay assessed in peripheral blood leukocytes of individuals residing in the vicinity of a mobile phone base station and comparing it to that in healthy controls (Gandhi et al., 2014). In epidemiological studies that assessed negative health effects of mobile phone base stations (seven studies explored the association between base station proximity and neurobehavioral effects (Navarro et al., 2003; Hutter et al., 2006;

Abdel-Rassoul et al., 2007; Berg-Beckhoff et al., 2009; Blettner et al., 2009; Gadzicka et al., 2006; Santini et al., 2002) and three investigated cancer (Wolf and Wolf, 2004; Havas, 2017; Levitt and Lai, 2010), 80% reported increased prevalence of adverse neurobehavioral symptoms or cancer in populations living at distances < 500 m from base stations (Navarro et al., 2003).

The literature also indicates that these effects may be cumulative based on i) mice exposed to low-intensity RFR became less reproductive and after five generations of exposure the mice were not able to produce offspring indicating intergenerational transfer of effects (Magras and Xenos, 1997); ii) DNA damage in cells after 24 h exposure to low-intensity RFR, which can lead to gene mutation that accumulates over time (Phillips et al., 1998) and iii) increased sensitivity to behavior-disruption experiments in rats (D'Andrea et al., 1986) and monkeys (de Lorge, 1984), iv) an increase in permeability of the blood-brain barrier in mice suggesting that a short-term, high-intensity exposure can produce the same effect as a long-term, low-intensity exposure (Persson et al., 1997). Studies on short-term exposure generally show no effects. For example, early studies saw no effect from

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short-term exposure, however, studies found effects after prolonged, repeated exposure in guinea pigs and rabbits (Takashima et al., 1979).

There are several studies showing the effect intensifies with reduced distance to the cell tower. The first (Santini et al., 2002) found increased symptoms and complaints the closer a person lived to a tower (Santini et al., 2002) and similar results were found in later studies (Navarro et al., 2003; Hutter et al., 2006; Abdel-Rassoul et al., 2007).

## 2. U.S. law unhelpful for preventing future liability

Current U.S. law has created a somewhat peculiar overriding federal preemption that precludes taking the “environmental effects” of RFR into consideration in cell tower siting (see Section 704 of The Telecommunications Act of 1996). The current, U.S. standards are based solely on thermal effects (which do not appear to be a problem) and thus do not mitigate against non-thermal effects (for which there is a growing litany of concern in the medical/scientific community). Due to the findings of many studies briefly summarized above many researchers argue for the revision of standard guidelines for public exposure to RFR from mobile phone base station antennas (Abdel-Rassoul et al., 2007; Hardell and Sage, 2008; Khurana et al., 2010). As Roda and Perry summarize (Roda and Perry, 2014), “... because scientific knowledge is incomplete, a precautionary approach is better suited to State obligations under international human rights law.” This is perhaps most forcefully concluded by the *BioInitiative Report* published by the BioInitiative Working Group, which is based on an international research and public policy initiative to give an overview of what is known of biological effects that occur at low-intensity electromagnetic fields exposure. This precautionary approach is gaining favor in Europe, but is less common in the U.S. American companies are therefore ill advised to simply follow “regulatory compliance” on this front, as there appears to be a clear cause for concern in the scientific/medical communities. If causation were to be proven through detailed studies, cellular phone companies would potentially be in position of future legal exposure for causing widespread human health problems and premature death. It is, therefore, in American companies’ best interest to act before government and regulation catches up with the science.

## 3. Current cell tower positioning

Current cell tower locations are chosen based on a “search ring” priority basis of geographic optimum for technical coverage of high concentration of wireless transmissions (e.g. users). This combination of technical parameters (e.g. geography) to enable coverage and dependable service and costs (e.g. positioning on mountaintops on accessibly by helicopter) is then weighed against and local regulations such as local zoning.

To overcome these challenges in urban areas cellphone companies often locate cellphone base stations at schools, because the monthly rental fee (~\$1500) is welcome income for economically-challenged school districts that have influence on local zoning. However, some jurisdictions have already prohibited the placement of cell phone towers near schools or hospitals because of the increased sensitivity of these populations, as in India. Other regions such as Europe (Roda and Perry, 2014) could follow a similar approach. Now even in North America, Canada’s Standing Committee on Health are considering more precautionary approaches to RFR.

## 4. Precautionary cell phone base station positioning

A review article of the health effects near base stations concluded that deployment of base stations should be kept as efficient as possible to minimize exposure of the public to RFR and should not be located less than 500 m from the population, and at a height of 50 m (Levitt and Lai, 2010). This potentially presents a serious challenge to cell phone company RF engineers. However, it is possible to obtain necessary

coverage while at the same time minimizing human exposure at the highest intensities. There are several first steps a cellular phone company can take to minimize human exposure particularly of the most vulnerable populations.

First, voluntarily restrictions can be made on the placement of cellular phone base stations within 500 m of schools and hospitals. This will synchronize base station deployment strategies between regions. This can be done by utilizing the existing hexagon planning map structure of an area with an overlay using an additional semi-automated process with a geographic information system (GIS) (Al-Sahly et al., 2018) such as the Geographic Resources Analysis Support System (GRASS) to identify any regions within 500 m of existing schools and hospitals. All hexagons with schools or hospitals are marked as unusable for RF engineer planning (e.g. colored red). This restriction only makes planning slightly more difficult, but does present a challenge in regions where schools were specifically targeted as base station locations in (e.g. Verizon deployments in the U.S.). Future work is needed to determine if the increased legal exposure warrants the cost of moving existing stations. However, the increased cost to locate future stations away from schools and hospitals should be minimal.

The second technical hurdle is more challenging. Ideally, all cell phone users would have coverage while minimizing the population density near cellular phone base stations (thus minimizing health impacts). This can be planned using GIS tools, freely-accessible U.S. Census data, parcel data and/or satellite images. The population density can be color coded for straightforward decision making for RF engineers. As a cellphone base station costs \$250–350,000 to install in the U.S., using a precautionary approach to potential future regulation can save substantial relocation fees.

The cell phone industry should also consider cell splitting, small cell deployment, beam and null steering antennae as possible technical means for reducing RF exposure. Moreover, more research on cognitive radio should also be conducted, so that the overall RF exposure is reduced. These measures will ultimately benefit the entire telecommunications industry, while potentially significantly reducing global RF pollution.

Finally, exposed companies should consider funding large-scale epidemiological studies with personal dosimeters for strict dose measurement and straight-forward tissue exposure. By quantifying the human medical threat themselves, more appropriate long-term planning can be made to minimize the risk of liability from unintended human harm due to cellular phone base station siting.

## Financial disclosure

The author owns stock in the American Tower Corporation.

## Declaration of competing interest

The author has no conflict of interest.

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OPEN

# Radiofrequency EMF irradiation effects on pre-B lymphocytes undergoing somatic recombination

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Intense electromagnetic fields (EMFs) induce DNA double stranded breaks (DSBs) in exposed lymphocytes. We study developing pre-B lymphocytes following V(D)J recombination at their Immunoglobulin light chain loci (*IgL*). Recombination physiologically induces DNA DSBs, and we tested if low doses of EMF irradiation affect this developmental stage. Recombining pre-B cells, were exposed for 48 h to low intensity EMFs (maximal radiative power density flux  $S$  of  $9.5 \mu\text{W}/\text{cm}^2$  and electric field intensity 3 V/m) from waves of frequencies ranging from 720 to 1224 MHz. Irradiated pre-B cells show decreased levels of recombination, reduction which is dependent upon the power dose and most remarkably upon the frequency of the applied EMF. Although 50% recombination reduction cannot be obtained even for an  $S$  of  $9.5 \mu\text{W}/\text{cm}^2$  in cells irradiated at 720 MHz, such an effect is reached in cells exposed to only  $0.45 \mu\text{W}/\text{cm}^2$  power with 950 and 1000 MHz waves. A maximal four-fold recombination reduction was measured in cells exposed to 1000 MHz waves with  $S$  from 0.2 to  $4.5 \mu\text{W}/\text{cm}^2$  displaying normal levels of  $\gamma\text{H2AX}$  phosphorylated histone. Our findings show that developing B cells exposure to low intensity EMFs can affect the levels of production and diversity of their antibodies repertoire.

Somatic or V(D)J recombination is the process that assembles in all jawed vertebrates the gene segments encoding the variable regions of the specific antigen immune receptors (T cell and Immunoglobulin IG) of the lymphoid T and B cells<sup>1</sup>. This process occurs in lymphocyte precursors, is mediated by RAG (recombination activating gene proteins) recombinase a heterotetrameric complex made of a dimer of RAG1 and two monomers of RAG2<sup>2,3</sup>. RAG1 a member of the DDE transposase/Integrase family is the key catalytic component of RAG. RAG binds specifically to recombination signal sequences (RSS) flanking germinal coding V, (D), J gene segments in the variable region at the IG and T cell receptor loci and catalyzes their rearrangement<sup>4</sup>. RAG recombination generates two DNA hairpins at the coding ends and two blunt double stranded DNA cuts at the signal ends. RAG maintains the paired cleaved ends in proximity and allows the ubiquitous set of non-homologous end-joining (NHEJ) DNA repair enzymes (Artemis, ATM, DNAPk, XRCC4, DNA Ligase IV) to resolve the hairpins and join the cleaved ends. For B and T lymphocytes recombination occurs at two stages during their differentiation<sup>5</sup>. We will discuss only the B lineage development in the bone marrow. First two rounds, D to J (in pre-pro stage) followed by V to DJ recombination (in late-pro stage) occur in pro-B cells at their Ig Heavy chain locus (*IgH*). Once *IgH* locus is rearranged, expressed *Igμ* together with a surrogate light chain comprising  $\lambda 5$  Vpre B proteins and two  $\text{I}\alpha$ ,  $\beta$  signaling subunits assemble the pre-B cell receptor (pre-BCR)<sup>6</sup>, which marks the large pre-B cell stage. Stromal bone marrow cells secreted interleukin IL-7 binds to their receptor (IL-7R), a signal which is transduced as pro-survival and proliferative<sup>7</sup>. First, IL-7R signals through Janus Kinase 3-(JAK-3)<sup>8</sup> phosphorylating and recruiting the signal transducer and activator of transcription 5A and B (STAT5A and B)<sup>9,10</sup> which stimulate transcription of *Ccnd3* encoding Cyclin D3<sup>11</sup> and of the B cell lymphoma 2(*bcl2*) gene<sup>12</sup>. Both Cyclin D3 and the anti-apoptotic BCL2 help pre-B cells through cell cycle G1 checkpoint allowing the replication of their DNA. Secondly, IL-7R signals in large pre-B cells through phosphoinositide 3-kinase (PI3K)<sup>13</sup> and protein Kinase B (AKT) phosphorylating the forkhead box O 1, 3 (FOXO1,3) transcription factors, modification which exports them from nuclei and targets the proteins for degradation<sup>14–16</sup>. FOXO1, 3 activate *e-rag* enhancer and *rag1*, 2 genes transcription<sup>14,17</sup>. In large pre-B cells IL-7R also signals via the nuclear factor kappa light chain

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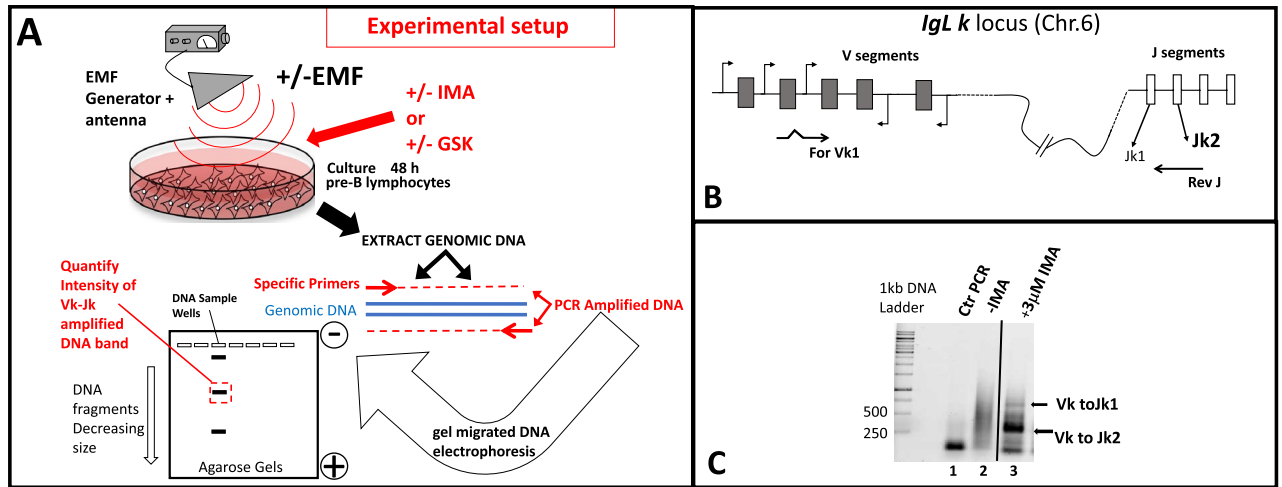
enhancer of activated B cells (NF- $\kappa$ B) stimulated by AKT phosphorylation of IKK $\alpha$  serine 23<sup>18</sup>. NF- $\kappa$ B activates Cyclin D4 kinase which targets FOXO1 for phosphorylation and repression<sup>19</sup>. By inhibiting FOXO1, or phosphorylating STAT-5, IL-7R signals are transiently downregulating RAG proteins in large pre-B cells. After four to five rounds of replication the large pre-B lymphocytes get under the influence of cell surface pre-BCR receptor aggregation and stimulation (in absence of a bonified ligand), a signal which antagonizes that of IL-7R, induces cell cycle arrest and transitions cells towards small pre-B stage<sup>20</sup>. Stimulation of pre-BCR cascades through RAS and extracellular signal- regulated kinase (ERK) upregulating the E2A transcription factor expression. E2A binds both Igk intronic and Igk 3' enhancers making the *Igk* light chain locus accessible for recombination<sup>21</sup>. Another effect of pre-BCR stimulation signals through spleen tyrosine kinase(SYK) and B cell-linker protein(BLNK) which together repress PI3K and AKT but stimulate mitogen activated p38 kinase which activates FOXO1 to express RAG<sup>13,20,22</sup>. Consequently, in small pre-B cells subsequent V to J rearrangements occur at *Ig L k* or  $\lambda$  light chain loci. Upon completion of a successful V to J recombined allele, the cell develops into naïve immature B cell, exposing IgM B cell receptors (BCR).

Interference of V(D)J recombination with other concurrent exogenous factors favoring DNA DSBs, like ionizing or EM irradiation can induce DNA damage which may lead to oncogenic translocations such as those described in acute lymphoblastic leukemia (ALL)<sup>23,24</sup>. Exposure of human blood lymphocytes from healthy donors to strong EMFs (2 h irradiation with sinusoidal pulses at  $4 \times 10^5$  V/m 50 Hz with a carrier wave of 10 Hz<sup>25</sup>) causes DNA DSBs and chromosomal lesions whose severity correlate with the intensity of the applied fields and the duration of exposure. However, less clear results come from studies with irradiated lymphocytes using low intensity, high radiofrequency(RF) EMFs (3 kHz–300 GHz)<sup>26</sup>. Most of these studies have assessed the levels of EMF inflicted DNA single and DSBs on lymphocytes using the microgel electrophoresis technique or 'comet assay', which detects breaks with a sensitivity limit of 50 strand events per diploid cell<sup>27</sup>. RF EM irradiation from cell phones was first studied by Phillips et al. in Molt-4 human lymphoblastoid cells exposed for 2–21 h to fields of 813.5 and 836.5 MHz with specific absorption rate (SAR) (2.4–26  $\mu$ W/g)<sup>28</sup>. Variable degree of DNA damage is reported, mainly induced by high SAR values waves (increased at 24 or 26  $\mu$ W/g and decreased at 2.4 or 2.6  $\mu$ W/g) and longer exposures (21 h versus 2 h). Another study by Mashevich et al.<sup>29</sup> reveals that continuous 72 h exposure of human peripheral blood lymphocytes to EMFs of 830 MHz waves, with SAR ranging from 1.6 to 8.8 W/kg lead to SAR dependent aneuploidy with specific abnormalities on chromosome 17. However, in vitro exposure of human blood lymphocytes for only 2 h to short pulses of 2450 MHz, at an average power of 5 mW/cm<sup>2</sup><sup>30</sup> showed no significant DNA damage as assessed by alkali comet assays. No signs of genotoxicity were found when total human blood leukocytes were in vitro exposed for 24 h either at a continuous or a pulsed-wave 1.9 GHz EMF with a SAR ranging between 0.1 and 10 W/Kg<sup>31</sup>. Absence of significant DNA damage response in human blood lymphocytes was also reported by a study by Stronati et al.<sup>32</sup> in which blood specimens were continuously exposed for 24 h at a Global System Mobile Communication generated EMF of 935 MHz with a SAR of 1 or 2 W/Kg<sup>32</sup>. Similar negative results with respect to EMF induced DNA damage was reported in a study by Hook et al.<sup>33</sup> with cultured Molt-4 human lymphoblasts exposed for 24 h to four types of frequency mobile network modulations around 815–850 MHz with SAR values ranging from 2.4 to 3.2 W/Kg<sup>33</sup>.

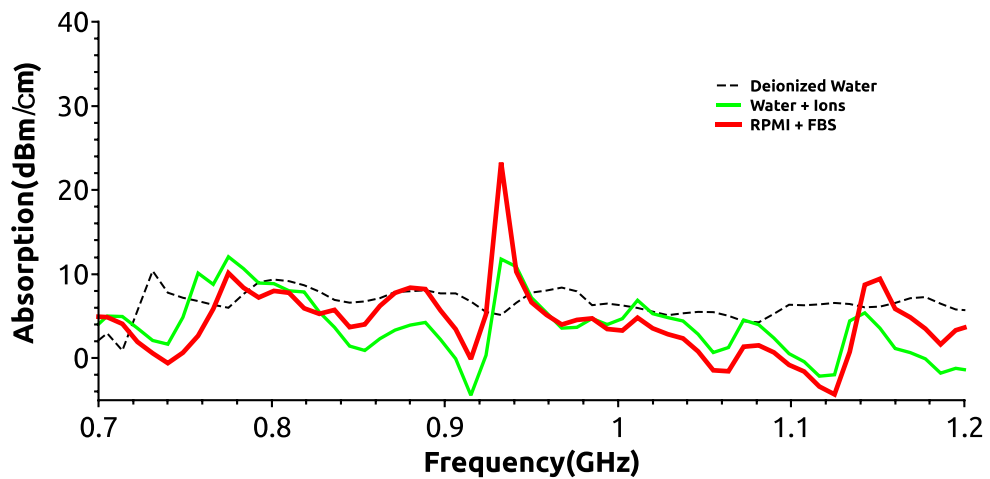
In our work we test the effects of in vitro irradiating V(D)J recombining pre-B cells with very low doses of RF EM waves. RAG stimulation is obtained either mimicking a pre-BCR stimulus with AKT inhibition, or with a stress inducible Abelson (Abl) kinase inhibitor response via STAT5 phosphorylation inhibition. For both stimuli, near 950–1000 MHz RF EMF cell irradiation, in the absence of detectable DNA DSBs, causes a four-fold reduction in recombination levels in exposed pre-Bs versus that assessed in non-irradiated cells.

## Results

**Design and specific experimental conditions used to assess *Ig k* locus rearrangements.** Our study tests how gene recombination levels are influenced by exposure to EMFs with distinct emitted frequencies and power levels (dose–response). In vitro grown vAbl transformed murine pre-B cells stimulated to recombine V(D)J are exposed to a broadband (0.8–3 GHz) emission antenna which broadcasts an EMF from a RF generator (Fig. 1A upper region). For all experiments we standardized our cellular growing conditions to control irradiation parameters (see Supplemental Material section S1 and Fig. 1Sa and b). RAG expression and V(D)J recombination can be induced in vAbl transformed pre-B cells(differentiating them in small pre-B cells) upon stimulation either with an Abl tyrosine kinase inhibitor imatinib(mesylate of imatinib)(IMA)<sup>34,35</sup>(Supplemental Material Fig. 1Sb growing dish wells 1, 2 and 3), or with an AKT inhibitor GSK-690693(GSK)<sup>19</sup>(wells 4, 5 and 6, Fig. 1Sb). Whereas IMA induces RAG by inhibiting vABL-1 tyrosine kinase via a stress-inducible GADD45 $\alpha$  action<sup>17,34,35</sup>, GSK acts as AKT inhibitor, reducing NF- $\kappa$ B and FOXO1 inhibitory phosphorylation (by CDK4) thus, mimicking a physiologic pre-BCR stimulation<sup>19</sup> (see Supplemental material section S2). Time course experiments with RAG induction in vAbl pre-B cells using both drugs show maximal RAG1 levels after 36 h of stimulation (see Supplemental material S2 and Fig. 2Sa and b). Using this finding, after 48 h post drug induction (to allow recombination), all synchronized cultured cells were harvested and their genomic DNA extracted. A previously described two-steps nested PCR (polymerase chain reaction) which assesses the recombination extent taking place at *Igk* kappa light chain locus (chromosome 6, locus schematic and primer positions shown in Fig. 1B), is templated with the equivalent genomic DNA extracted from  $2 \times 10^6$  cells from each tested culture set<sup>36,37</sup>. In the absence of V(D)J recombination (control reactions with no stimulation Fig. 1C lane 2) the variable region V and J segments in germline configuration are too far apart on the chromosome to yield appropriate amplification products. The PCR amplification products obtained only from recombined templates (Fig. 1C lane 3) are separated after electrophoretic migration on 1.5% agarose gels and visualized after fluorescent staining with SYBR green (schematic lower drawing Fig. 1A, and gel scan Fig. 1C). This typical nested PCR reaction reports *k* locus recombination events with two detectable products; the predominant one Vk-Jk2 of 280 bp (95%) and Vk-Jk1 of

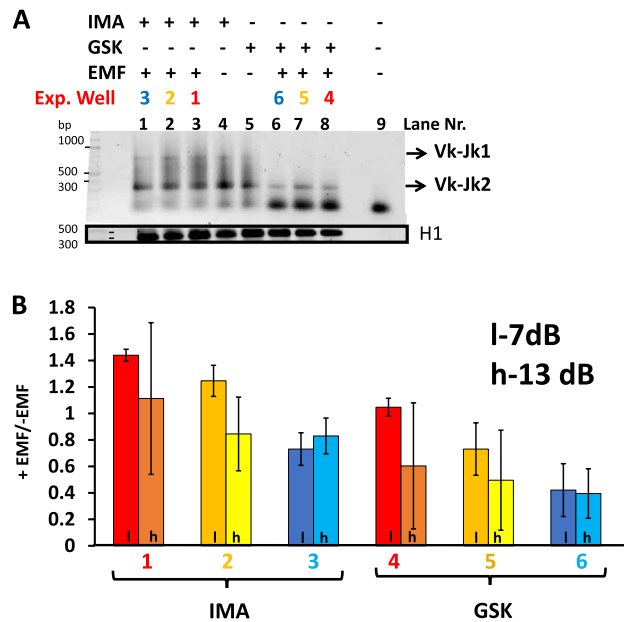


**Figure 1.** (A) Schematic depiction of the flow chart of the experimental design. Murine A-70 vAbl pre-B cells grown with or without exposure to Electromagnetic field influence(EMF), in the absence or presence of RAG stimulation either by Mesylate of Imatinib (IMA) or by GSK-690693(GSK), are harvested and their genomic DNA subjected to a two-steps PCR recombination assay identifying Vk to Jk rearrangements at their *IgL kappa* loci. The electrophoretically separated recombination products (Vk-Jk) are quantified by densitometry to assess the extent of locus rearrangements influenced by EMF. (B) Schematic configuration of *IgL k kappa* locus on Mouse chromosome 6, and the positioning of the primers used in the assay. (C) PCR reactions electrophoretically separated in agarose gel stained with Sybr green identify the recombined products (arrows show Vk to Jk1 and Vk to Jk2) in lane 3 versus, control reactions lane 1 without genomic DNA, lane 2 templating genomic DNA from uninduced cells (in germline configuration). Such recombination amplified reactions are then used for densitometry quantifications. The entire gel from which (C) was cropped displaying amplifications (Vk to Jk response) from cells treated with a wide range of increasing IMA concentrations , is shown in Supplemental Material Fig. 3Sa.



**Figure 2.** Absorption spectra measurements of filtered deionized water (dashed thin black line), tap water (green thick line Water + Ions) and RPMI cell culture medium with 10% fetal calf serum (FBS)(red thick line RPMI + FBS) All measurements were done using a Keysight-AGILENT-HP N9935A spectrum analyzer as described in “Methods” section.

700 bp (5%)<sup>36</sup> (Fig. 1C lane 3). Densitometric quantifications of the DNA Vk-Jk2 recombination products allow us to assess the EMF influence on recombination (Fig. 1A lower drawing). A dose–response (recombination) effect obtained with increasing IMA concentrations in 48 h stimulated pre-B cells is shown in supplementary Fig. 3Sa, gel and quantified data from three such experiments shown in Fig. 3Sb histograms. The lowest drug concentration (3 μM for IMA and of 10 μM for GSK,) for which maximal recombination effects are obtained, is used for each drug in our irradiation assays. For linear range quantifications of the image scans each reaction uses genomic DNA template at least at three distinct dilutions from the cellular extraction stock solution and the final result may be reported as an average of the three quantified products values corrected by the histone H1 band intensity of the corresponding sample. In Supplemental material in Fig. 3Sc an 3Sd a set of nested PCR



**Figure 3.** A two steps PCR recombination assay is used to identify Vk to Jk1 or Vk to Jk2 rearrangements from pre-B cells upon RAG induction with Imatinib or GSK. **(A)** A Sybr Green stained 1.5% Agarose TBE gel in which the recombination PCR reactions templated with initial 1:5 dilutions of genomic DNA extracted from each distinct cell treatment lot ( $2 \times 10^6$  cells) are electrophoretically resolved. The cells were either unexposed (gel reaction lanes 4 and 5) or subjected for 48 h to 1 GHz EMF irradiation (lanes 1 to 3 and 6 to 8) with the generator setting at 13 dBm. The color code designating the positions of exposed EMF (exp.Well) wells in the dish is the same with the one used in supplementary Fig. 1Sb. Last lane (9) of the gel, -DNA control reaction. The bottom black box (cropped from a distinct gel) displays Histone H1 PCR reactions templated with the same amount of genomic DNA as the recombination reactions above (control genomic DNA). **(B)** Identified Vk to Jk2 recombined products were quantified from scanned gels corresponding to PCR reactions from cells +/- Irradiation and the calculated ratios of band intensities expressed +EMF/-EMF (irradiated/nonexposed) for each well (color code consistent with that shown in Fig. 1S). The histograms represent the average values of three independent quantified experiments. EMF-Electromagnetic Field, Recombination pharmacological stimuli (Imatinib, IMA) versus (GSK-690693, GSK). H1, histone H1 control reaction PCR reactions. Darker font histograms correspond to lower 7 dBm (l) and brighter to higher 13 dBm (h) generator power settings.

reactions templated with serial dilutions of input genomic DNA from IMA stimulated cells, followed by quantification of the signal are shown to illustrate that the assay responds linearly in its amplified Vk-Jk2 band intensity.

**EM wave absorption spectrum of the cell culture medium.** We measured how the EM waves with frequencies ranging from 700 to 1224 MHz are absorbed by the fetal bovine calf serum supplemented cell culture medium (RPMI + FBS in Fig. 2) in which the pre-B cells are cultured. For comparison only absorption measurements were also performed for deionized water (conductivity  $< 5 \mu\text{S}/\text{cm}$ ), and for ions containing unfiltered tap water samples (see Methods Water + ions, Fig. 2). The measurements were done using a setup in which an emission and a reception horn antenna were spaced 1 m apart with the liquid sample container positioned in the vicinity (1 cm) of the later (see Supplemental material Fig. 4S). The emission antenna was connected to a generator and signals from the receiver antenna were collected and recorded by a standard spectrum analyzer. In Fig. 2 are presented the background corrected absorption spectra per 1 cm width of each liquid sample measured. A well-defined absorption peak is observed at 938 MHz for the RPMI + FBS medium sample which is twice as large as the others measured at this frequency. All samples have similar absorption values for the rest of the tested spectral frequencies. This finding is important since the range of frequencies (720 MHz, 850 MHz, 950 MHz, 1 GHz and 1.2 GHz) to be used for cell irradiation centers our window of exposure between 950 MHz and 1 GHz, proximal to the maximal culture medium absorption peak.

To test how the cell growing medium affects the electric intensity of the exposing fields, EMF electric flux density (D displacement) measurements were made inside the incubator for each mentioned frequency, in the absence or presence of culture medium in the culture plate (Supplemental Material S3 and Fig. 5S). Values greater than one of the  $D_m/D_{\text{air\_inc}}$  (1.8–1.95) ratios measured between 750 and 1000 MHz (Supplemental material section S3 and Fig. 5Sc) show in this range, the complete RPMI + FBS cell growing medium selectively potentiates the developed fields.

**EMF irradiation effects on V(D)J recombination in v-Abl pre-B cells.** Murine vAbl pre-B cells were grown under normal conditions or stimulated either with 3  $\mu\text{M}$  IMA or with 10  $\mu\text{M}$  GSK in the presence/absence of an antenna which emits a generator controlled EMF from waves of 720 MHz, 850 MHz, 950 MHz,

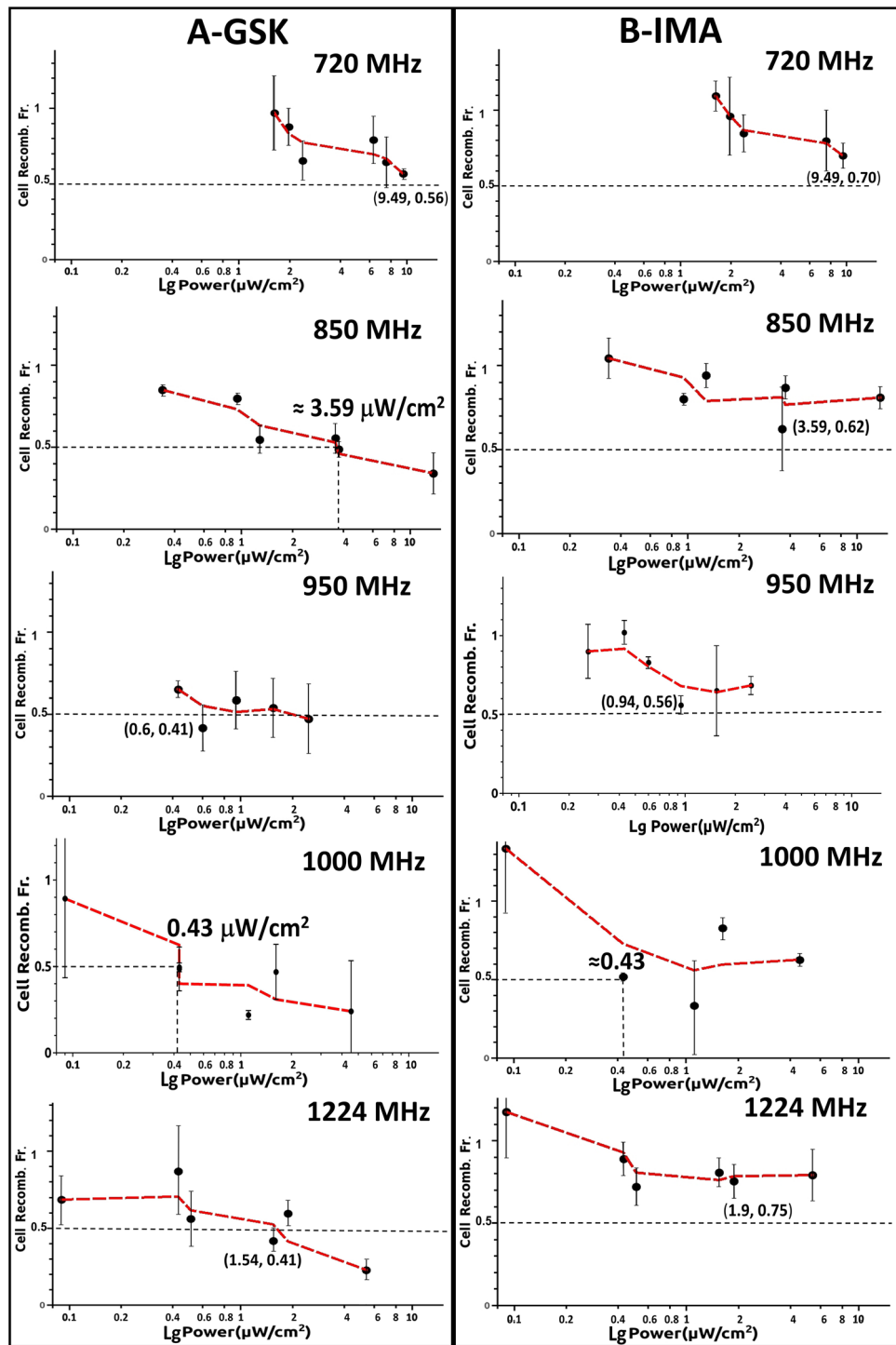
1 GHz, 1.224 GHz each with 7 or 13 dBm output power setting. For all exposures, the antenna was held at 2.4 cm above the composite 6 wells plate as depicted in Supplemental material Fig. 1Sb (lower profile drawing) consistently keeping it in the same location with respect to the incubator walls (Supplemental material S1 and Fig. 1S). Cells were grown +/- EMF constant continuous exposure for 48 h with +/- IMA or +/- GSK. In Fig. 3A is shown a gel with resolved reactions either from nonexposed cells (lanes 4 and 5) or from cells continuously subjected for 48 h to the influence of 1 GHz fields (gel for generator set at 13dBm-h), with both RAG induction treatments (plate Exp. wells IMA 1, 2, 3 and GSK- 4, 5, 6 with color code shown in Supplemental material Fig. 1Sb). Visually one can see, a reduction of Vk to Jk2 recombination products obtained in reactions from irradiated cells versus those from similarly drug induced, non-irradiated cells (see Fig. 3A compare lane 4 non-irradiated to reactions in lanes 1–3 exposed for IMA, and lane 5 unexposed to lanes 6–8 from irradiated GSK stimulated cells). The irradiating effects are most pronounced in the plate wells closest to the actively emitting antenna elements ( $\lambda/2$  for 1 GHz waves use as main element the 15 cm one located near wells 3 and 6 (Supplemental material Fig. 1Sb) hence, recombination reduction for plate Exp. wells 3, 6 > 2, 5 > 1, 4 or correspondingly gel lanes 1, 6 > 2, 7 > 3, 8). The value of the calculated ratios between recombination Vk-Jk2 PCR band intensities obtained from irradiated/non-irradiated(+EMF/-EMF) cells for all tissue culture wells are shown as histograms in Fig. 3B. Values less than one show specific Vk-Jk2 recombination reduction associated with EMF irradiation.

Similar experiments were performed with EMF exposures at 720, 850, 950, 1000 and 1224 MHz (each frequency centers on a different antenna element), generator setting either at 7 dBm or 13 dBm. To display a wider palette of EMF dose exposure values we summed up the data from all of the wells in Fig. 4 which displays cell Vk-Jk2 recombination Fractions(+EMF/-EMF -ordinates), against logarithm of measured irradiating power flux density S values ( $\mu\text{W}/\text{cm}^2$ -abscissas) at each location. Each row of the two panels is for a distinct frequency with panels for each drug located on the same column: Fig. 4A(GSK- left) and B(IMA- right). Consistently all diagrams show power dependent reduction in cellular Vk-Jk2 recombination. S values into the emitting antenna were calculated from antenna recorded voltages, circuit impedance, and antenna constructive elements dimensions and reflect S in the air inside incubator, surrounding the involved culture well. In each panel with dotted black lines we pointed the EMF power dose required to induce a two-fold Vk-Jk2 recombination reduction from that of the non-irradiated lot (+EMF/-EMF 50% reduction shown as 0.5 ratio for Vk-Jk2, Cellular Recombination Fraction). In Fig. 4 when 50% recombination reduction (exposed versus non-irradiated cells) is not reached, the minimal recombination ratios obtained and their inducing S levels are shown in parenthesis. The most remarkable finding of our study is that even for such a small window of frequencies (between 720 and 1224 MHz), the power dose-response effect is dramatically influenced by the frequency of the irradiating EMF. If at 720 MHz one reaches a 0.56/0.70 maximal recombination reduction for 9.49  $\mu\text{W}/\text{cm}^2$  exposure, similar reduction in recombination effects are obtained at 950 MHz and 1 GHz with only 1/15th respectively 1/20th (0.63 or 0.43  $\mu\text{W}/\text{cm}^2$ ) the power used at 720 MHz. The power dose-cell recombination response curves at 950 MHz and 1 GHz EMFs show by far the most accentuated measured effects (for both drugs). For GSK at 1 GHz irradiation, an almost four-fold decrease in V(D)J recombination (from 0.90 to 0.22) is observed over a moderate increase in S exposure from 0.1 to 4.53  $\mu\text{W}/\text{cm}^2$  (see second from the bottom panel in Fig. 4A GSK 1000 MHz). Both curves in Fig. 4 for 1 GHz display an abrupt recombination decrease at a small increase in S (0.25–1  $\mu\text{W}/\text{cm}^2$ ) after which the cellular effect plateaus out over a larger window of higher exposure power S values (1–4.5  $\mu\text{W}/\text{cm}^2$ ). To emphasize the influence of EMF frequency Table 1 shows how recombination fractions (+EMF/-EMF) vary at a relatively constant  $\approx 1.5 \mu\text{W}/\text{cm}^2$  irradiating power flux density S exposure level for all tested EMF frequencies. At this small irradiating power no effect is detectable at 720 MHz, whereas at 950 MHz a two-fold recombination reduction is measured reaching almost three-fold recombination inhibition at 1 GHz.

To circumvent the cellular growing medium polarization effects (which significantly change polarity at 720 MHz and above 1100 MHz, Supplemental material 3S and Fig. 5Sc) or its enhanced wave absorption at 938 MHz (Fig. 2), we intentionally represented in Fig. 5 the recombination fractions for two constant electric field intensity E exposure values, measured inside the medium; one of 0.4 V/m (Fig. 5A.) and the other of 0.6 V/m (Fig. 5B). The approximative intensity of the emitted electric field was calculated in the cell medium from the measured electric flux density ( $D_m$  displacement) values<sup>38</sup> described in the previous section, and averaged for the central plate well. For both E values and both pharmacological stimuli (IMA-red and GSK-blue) the most accentuated plots concavities (maximal irradiation induced recombination reduction effect), correspond to 950–1000 MHz. At both E values represented in Fig. 5 the recombination ratios are unaffected by EMFs at 720 MHz. In contrast, at 1000 MHz, a two-fold reduction is observed for the 0.4 V/m EMF intensity, and a three (IMA) to four-fold (GSK) decrease is measured at the stronger 0.6 V/m field exposure. The electric fields dose exposures -recombination reduction effects in Fig. 5 and those reported for EMFs power dose exposures in Fig. 4 are qualitatively similar. These data strongly suggest that exposure even to very low irradiation doses from specific 900–1000 MHz radiofrequency waves dramatically affect in irradiated pre-B cells the efficiency of V(D)J recombination at their Ig kappa locus.

### Histone H2AX phosphorylation shows no detectable DNA DSB damage cell response in EMF exposed pre-B cells.

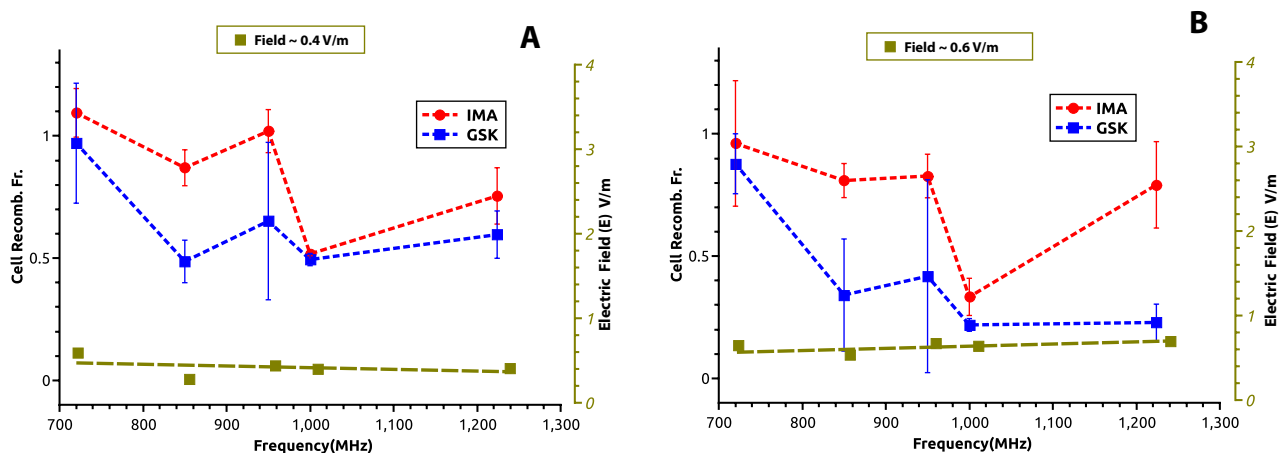
We asked whether the observed EMF irradiating effects on V(D)J recombination are due to DNA damage and presence of unrepaired DSBs. Impairment of DNA integrity can be assessed by the extent with which irradiation induces H2AX histone phosphorylation ( $\gamma\text{H2AX}$ ), a process associated with DNA DSBs and their intranuclear repair. The nuclear  $\gamma\text{H2AX}$  repair foci are the fairest indication that the NHEJ DNA repair machinery acts properly in these cells repairing DSBs caused by any DNA lesion-causing agent<sup>39,40</sup>. We grew cells under similar stimulation (+/- IMA, +/- GSK) and +/- EMF irradiation conditions (7 dBm or 13 dBm generator power settings at 950 MHz) with those described above but instead of extracting DNA, the harvested cells were fixed and doubly stained: (a) with Hoechst 33342 dye (for nuclear total DNA staining in



**Figure 4.** The EMF power dose-cell recombination response curves at 720, 850, 950, 1000 and 1224 MHz for both types of pharmacological agents stimulating RAG expression (A) (GSK-690693, GSK), and (B) (Imatinib, IMA). Cell Recomb. Fr. expresses the ratio values of measured Vk-Jk2 recombination quantified from cells grown in +EMF/-EMF (irradiated/non-exposed) conditions. Bottom abscissa displays logarithm of S power flux density values (Power  $\mu\text{W}/\text{cm}^2$ ) measured around the emitting antenna inside the CO<sub>2</sub> 5 vol%, and 95% water humidity incubator air conditions, expressed as a single range in all panels(logarithmic scale). The black dotted line denote a level of EMF induced two-fold recombination reduction (Cell recomb. Fr.=0.5), whereas when this level is not reached in the experiment the coordinates of the lowest obtained Cell Recomb. Fr. are given. The red dotted line connecting markers is just a Moving Window Average line which accounts for the average between successive data points displaying the trend of data variation. The error bars represent standard deviation (SD) values from three independent experiments.

Fraction recombination +EMF/-EMF (EMF at $S \approx 1.5 \mu\text{W}/\text{cm}^2$ )		
Frequency (MHz)	Response stimulus	
	GSK	IMA
720	$0.97 \pm 0.2$	$1.09 \pm 0.1$
850	$0.56 \pm 0.1$	$0.8 \pm 0.1$
950	$0.53 \pm 0.2$	$0.65 \pm 0.3$
1000	$0.38 \pm 0.1$	$0.46 \pm 0.3$
1224	$0.41 \pm 0.1$	$0.8 \pm 0.1$

**Table 1.** Lists the measured cell recombination fraction (+EMF/-EMF) at a relative constant power flux density  $S$  value of  $1.5 \mu\text{W}/\text{cm}^2$  for all tested frequencies.

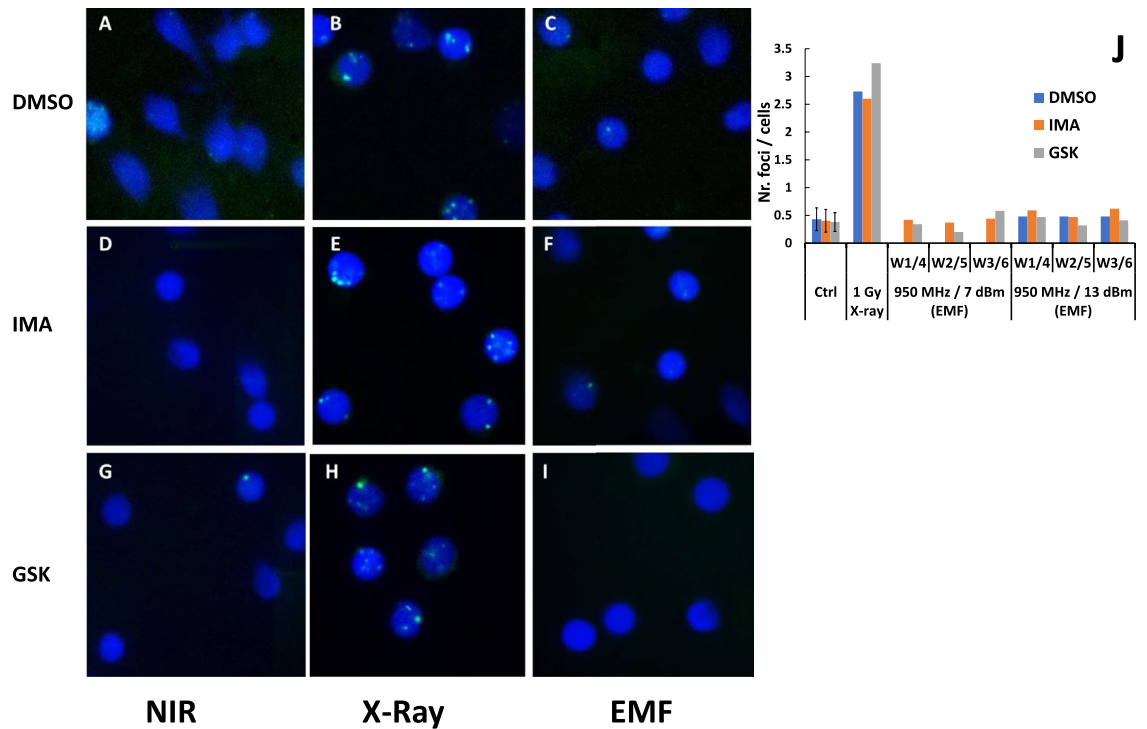


**Figure 5.** Variation of cell recombination fraction (+EMF/-EMF) with the field irradiation frequency shown in each panel for a constant receiver EMF electric intensity field  $E$  calculated in the cell culture medium. (A) EMF electric field intensity  $E$  0.4 V/m, (B) EMF electric field intensity  $E$  0.6 V/m. The pharmacological agents stimulating RAG expression GSK-690693, GSK-blue, and Imatinib, IMA-red. The pale green dotted line shows the relative constant distribution of measured electric field as a function of frequency. The error bars represent standard deviation (SD) values from three independent experiments.

blue) and (b) immunofluorescently with anti  $\gamma\text{H2AX}$  antibodies yielding a Cy2 green fluorescence which identifies DNA DSBs repairing  $\gamma\text{H2AX}$  foci<sup>40</sup>(see Methods). As a DNA DSBs control an extra lot of cells were either noninduced or similarly drug treated but instead of EMF they were subjected to a quick 1 Gy, X ray irradiation dose prior to their harvest. Nine immunofluorescent images are shown in Fig. 6 A-I where blue contours show the cell nuclei and the green dots the DNA DSBs repairing  $\gamma\text{H2AX}$  foci from: cells treated with +/- DMSO solvent control, +/- GSK, +/- IMA, +/- EMF set at 950 MHz, 7dBm exposure and the control lot of cells exposed to 1 Gy X ray. Such foci were also counted and their number reported per cell to a number of total 100 counted cells gathered from more than twenty successive field views for each experimental lot (shown as histograms in Fig. 6J) for both 7 dBm and 13 dBm generator power settings). 1 Gy dose X ray irradiated cells are shown in Fig. 6B control with DMSO solvent, E with IMA, H with GSK and in 6 J the corresponding foci/cell counted histograms. All images (Fig. 6B,E,H) and the quantified histograms from X ray irradiated cells show similar and considerable DNA DSB lesions with consequent accumulation of  $\gamma\text{H2AX}$  repair foci, regardless of the chemical stimulus used. On the contrary, the long 48 h EMF exposure experiments do not show signs of detectable unrepaired DNA DSB damage (Fig. 6C DMSO solvent, F with IMA and I with GSK, and counted foci/cells in Fig. 6J), above the background level of non-irradiated control cultures (Fig. 6A,D,G and ctrl. histograms in Fig. 6J). Exposing for 48 h cells to EMF, regardless of drug treatment, does not seem to inflict significant/ cumulative unrepaired DNA DSB lesions, (unlike those caused even by mild quick irradiation with 1 Gy dose of X rays). Only such DNA injuries could have caused a detectable accumulation of repairing  $\gamma\text{H2AX}$  foci at the time of their harvest. Indirectly, these results suggest that the significant EMF induced reduction in pre-B cells recombination reported in Figs. 3B, 4, 5 and Table 1 is probably not caused by an enhanced level of accumulated unrepaired DNA DSBs.

## Discussion

V(D)J recombination the central process in lymphocyte development physiologically generates DNA DSBs during its course, when cells become susceptible to external sources of DNA damage<sup>5</sup>. Our work tests how pre-B lymphocytes exposure to low dose EMFs of frequencies ranging from 720 MHz to 1.2 GHz, used in utilitarian purpose telecommunication, affects the efficiency of their *Igk* loci rearrangements. First, we established a setup



**Figure 6.** (A–I) Immunofluorescence detection of  $\gamma$ H2AX phosphorylated histone foci in pre-B cells exposed to EMF or X ray irradiation. The  $\gamma$  H2AX foci are shown in bright green— $\gamma$ -H2AX, whereas DNA staining is in blue – nuclei. (A) Control solvent (DMSO) treated cells Nonirradiated (NIR); (B) Control (DMSO) treated 1 Gy X-ray irradiated cells(X-Ray); (C) Control (DMSO)treated EMF exposed (waves at 950 MHz, with emission generator power setting set at 7 dBm-EMF); (D) IMA treated NIR; (E) IMA, X-ray; (F) IMA, EMF; (G) GSK, NIR; (H) GSK, X-ray; (I) GSK, EMF. (J) Number of counted foci per /cell represented as histograms. W1/4 refers to growing plate wells 1 and 4, W2/5 wells 2 and 5 and W3/6 wells 3 and 6 equivalent positions with respect to which cells were EMF irradiated, with generator power setting set at 7 dBm and 13 dBm. GSK is cells treatment with 10  $\mu$ M GSK-690693, IMA their treatment with 3  $\mu$ M Imatinib. ANDOR camera assisted by IQ Live Cell Imaging software and foci analysis with Imaris for Cell Biologists software (both from OXFORD Instruments).

to control the EMF developed inside the cellular growing medium in a typical cell culture incubator. Cultured pre-B cells synchronously recombining V(D)J were EMF exposed during a 48 h window, which starts with RAG expression and ends with the NHEJ DSBs DNA repair<sup>5</sup>. A nested PCR assay is then used to study the cellular EMF irradiation gene effects.

The measured data in Figs. 3B, 4, 5 and Table 1 consistently show, EMFs cause a dose dependent reduction in V(D)J recombination in the irradiated pre-B cells, with similar effects for both RAG inducing stimuli (IMA or GSK) used. The magnitude of effects is tightly determined by the EMF frequency. A two-fold reduction in V<sub>k</sub>-J<sub>k2</sub> recombination at *Igk* locus narrowly can be obtained by an emitted S power value of 9.49  $\mu$ W/cm<sup>2</sup> at 720 MHz (Cell Recomb. Fr. 0.56 for GSK and 0.7 for IMA), whereas this effect may be achieved by a field developing one twentieth of that S dose at 1000 MHz (0.5 Cell Recomb. Fr. for both drugs at 0.43  $\mu$ W/cm<sup>2</sup>) (Fig. 4). The recombination reduction although observed for all EMFs tested, seems to be maximal for 950 and 1000 MHz waves, a small domain where the serum containing cell culture medium displays maximal EMF absorbance (Fig. 2), and augments by its molecular polarization the EMF electric intensity (supplementary Fig. 5Sc). We measured EMF local antenna emissive S values only in the incubator air surrounding the cell culture plate. Despite this limitation we measured and calculated the average irradiated electric field intensity E, inside the culture medium. The maximal effects were measured at 950 and 1000 MHz, where *Igk* recombination levels for an EMF of E 0.4 V/m are only half (Fig. 5A), or for one of 0.6 V/m E a quarter of those reported for same E values at 720 MHz (Fig. 5B). E dose effects parallel the frequency dependency described for the antenna emitted power dose S. However, the cell medium electric properties mentioned above, (increased absorbance and polarization between 900 and 1000 MHz), cannot account for the frequency results shown in Fig. 5 for irradiations at constant electric field E values. Besides such intrinsic medium properties there must be also a major EMF frequency direct influence on the cellular components linked to recombination.

Various wireless network service providers use for mobile phone communication frequencies ranging between 700 and 2100 MHz. At 1 cm distance, during outgoing calls the measured emitted field E intensities vary with  $\pm$  5–15% from the 41.25 V/m (recommended ICNIRP value) with cell phone models, whereas their maximum output recorded power levels for a GSM1800 net varies between 0.25 and 1 W<sup>41,42</sup>.

We assessed if the low dose 48 h EMF irradiations cause DNA DSBs and detectable  $\gamma$ H2AX repair foci in exposed cells. From the levels of detected  $\gamma$ H2AX repair foci of the EMF irradiated pre-B cells we could not reveal in exposed cells above background DNA DSBs repair activity (Fig. 6 compare panel A with C, G with I, and histograms in Fig. 6j). Using chromatin immunoprecipitation (ChIP) Savic et al.<sup>43</sup> show considerable  $\gamma$ H2AX accumulation near Jk5 in IgK locus after 24 h post STI571 (Imatinib) treatment of pre-Bs, but a dramatic more than two-fold decrease in  $\gamma$ H2AX detection as cells were kept from 24 to 48 h post STI571 treatment<sup>43</sup>. We could not detect above background  $\gamma$ H2AX foci levels in IMA or GSK treated cells after 48 h culture growth. This could be due either to a considerable post RAG DSBs repair recovery, or to a reduced sensitivity of our immunofluorescence assay (less sensitive than ChIP in detecting  $\gamma$ H2AX). The onset of DNA DSBs either prior to or during pre-B cells maturation inhibits *rag1*, 2 transcription<sup>44</sup> and reduces the levels of *Igk* locus rearrangement events<sup>45</sup>. These cellular stress effects are caused by ataxia teleangiectasia mutated (ATM) kinase either via NF- $\kappa$ B, FOXO1 signaling<sup>44,46</sup> or via GADD45a inhibition<sup>45</sup>, both pathways directly targeting *rag* genes transcription levels. If very few EMF induced DSBs (below those detectable by  $\gamma$ H2AX foci assay), or breaks already repaired before our cell harvests could have reduced RAG expression in our experiments (via ATM kinase) this could explain our observed reduced recombination effects. We used in our experiments two RAG induction stimuli, IMA sensitive to ATM kinase via GADD45a inhibition<sup>44–46</sup> and the second GSK690693 AKT-inhibitor insensitive to this signaling pathway<sup>19</sup>. If very few EMF induced DNA DSBs would have reduced RAG expression prior to, or during drug action, one would have expected experiments to show a more accentuated recombination reduction for IMA than that obtained for GSK treatment. Instead, the experimental data in Figs. 3B, 4, 5 and Table 1 show for both drugs very similar EMF induced reduction of *Igk* loci rearrangement levels (if not even slightly more pronounced reduction for GSK). IMA although a more potent RAG inducer than GSK has the disadvantage that post recombination blocks cells in Go phase preventing further their division<sup>36,47–49</sup>. On the contrary, the AKT inhibitor GSK-690693 not only is a weaker RAG induction stimulus (closer to the one physiologically occurring in small pre-B cells)<sup>50,51</sup> but also enables cells to divide prior to and after *Igk* loci rearrangements and protect their progress to the next stage of development<sup>19</sup>. Because our PCR assay intentionally uses the amount of templating genomic DNA from the same number of 2 millions harvested cells, replication would have “diminished the EMF recombination reduction” in GSK treated cells in contrast to those incubated with IMA (the later, on the contrary, “freezes” the EMF effect on BCL2 maintained survivors). As pointed earlier, in treated cells, both drugs show very similar EMF induced reduction of rearrangements in treated cells. Although we cannot fully refute that the observed EMF recombination effects may have been caused in irradiated pre-B cells by undetectable DNA DSBs via ATM, the line of evidence gathered from our experiments in Figs. 4, 5, 6 and the arguments presented above for the comparative IMA/GSK treatments make this mechanism a less likely candidate for their account.

Indirectly our work addresses the longstanding question of how innocuous low dose EMF irradiation from our telecommunication devices may be and whether it may affect the immunity of our organisms. It remains only to our speculation to extend the observed recombination effects induced by small EMFs from an in vitro culture system to the in vivo situation on the ability of irradiated B cells to elicit an unaltered antibody response to antigen challenge.

## Methods

**Materials.** DNA oligonucleotides were purchased from Life Technologies and IDT DNA: Vk degenerate primer 5' GCTGCAGSTTCAGTGGCAGTGGCAGTGGRTCWGGGRAC 3' where S is G or C, R is A or G, W is T or A, Jk2-1 primer 5' CAAAACCCTCCCTAGGTAGACAATTATCCCTC 3' and Jk2-2 primer 5' GGACAG TTTTCCCTCCTTAACACCTGATCTG 3'. For Histone H1 gene control amplifications the following primers were used: H1fw 5' GGCTGCTATCCAGGCAGAGAAGAACCG 3', H1rv: 5' GCTTTGGAGGCGCCTTCT TGGGCTTG 3'.

*Murine pre B cells* transformed with Abelson virus (v-Abl preB, A70 line, that harbor a E $\mu$ -Bcl2 transgene) were a kind gift from Barry Sleckman Duke University<sup>47</sup>. The cells were maintained in RPMI 1640 medium, supplemented with 10% FBS (both from GIBCO), 50  $\mu$ M 2-mercaptoetanol and induced at  $0.5 \times 10^6$  cells/ml density either with 3  $\mu$ M Imatinib Mesylate (IMA) (SIGMA-ALDRICH) or with 10  $\mu$ M GSK-690693 (GSK) (GLAXOSMITHKLINE, SELLECK-chem) in solutions with 0.1% DMSO. After 48 h the cells were collected and analyzed using the nested PCR described below.

*Pre-B Cells irradiation* was performed with a 1 Hz–1.224 GHz, 13 dBm radiofrequency generator (Hameg Instruments 1 Hz–1.2 GHz programmable synthesizer HM8134-3, used throughout our study as emission generator) using a broadband irradiating 800 MHz–3 GHz LTE ATK-LOG ALP logarithmic antenna, in a regular CO<sub>2</sub> incubator (SANYO Electric Co. MCO-17AIC), with CO<sub>2</sub> 5 vol. %, and 95% purified water humidity. Cells were grown at 37 °C in 5 ml medium in standard six flat bottom wells (16.8 ml capacity) polystyrene lidded plates (Corning Costar), which were always positioned in the same place with respect to the incubator walls (in the center of the incubator, see Supplemental material Fig. 1Sa) and the emission antenna (antenna central guiding label positioned midway between wells 3 and 6 at 2.4 cm above the mid plane of the plate, see supplementary Fig. 1S). Two parallel sets of experiments were performed with wells 1, 2, 3 containing cells stimulated with 3  $\mu$ M IMA, whereas wells 4, 5 and 6 cells were stimulated with 10  $\mu$ M GSK (Fig. 1SB).

**Two steps nested PCR reactions for K locus recombination.** Template DNA was prepared for PCR using a modified technique developed by Schlissel<sup>37</sup>. Pre-B A-70 v-Abl cells were harvested after 48 h incubation with IMA<sup>36,47</sup>, GSK<sup>19</sup> or unstimulated. Cultured cells ( $2 \times 10^6$ –2 millions) were pelleted for 15 s in a microfuge, washed once in PBS (phosphate saline buffer pH 7.2), resuspended in 200  $\mu$ l PCR lysis buffer (10 mM Tris pH 8.4, 2.5 mM MgCl<sub>2</sub>, 50 mM KCl, 200  $\mu$ g/ml gelatin, 0.45% NP40, 0.45% Tween-20 (CALBIOCHEM), and 60  $\mu$ g/ml Proteinase K (Boehringer), and incubated at 56 °C for 3 h followed by 15 min at 95 °C. Dilution of templates

was done with PCR lysis buffer without Proteinase K. Two successive PCR amplifications were done in a final volume of 50  $\mu$ l containing 2 to 5  $\mu$ l template DNA, 10 mM Tris-HCl (pH 8.4; at room temperature), 50 mM KCl, 2.5 mM MgCl<sub>2</sub>, 200  $\mu$ g/ml gelatin, 0.2 mM of all four dNTPs (all from ThermoFisher scientific), each oligonucleotide primer at 0.4  $\mu$ M (20 pmol each primer per reaction), and 1 U TAQ DNA polymerase (GoTaq PROMEGA) in nested reactions. First step PCR reactions for 25 cycles use Vk, and Jk2-1 primers. In the second step various dilutions (from 4  $\mu$ l 1:100 dilution of first PCR to 0.5  $\mu$ l of the first undiluted PCR) are individually used to template the second PCR reactions to which Vk and Jk2-2 primers are added and an additional round of 30 cycles amplification is performed. Cycling steps were: an initial 1 min denaturation at 94 °C, then repeated cycles each, 30 s at 94 °C, 0.5 min annealing at 50 °C, and 1.5 min polymerization at 72 °C. A final additional 5 min extension step was performed at 72 °C<sup>36,37</sup>. PCR products were resolved on 1.5% agarose gel, stained either with ethidium bromide or Sybr Green (THERMOFISHER scientific) and visualized using the PharosFX system (BIORAD). The bands intensities were quantified using QuantityOne software.

**Kappa locus amplification products analysis.** Each Vk-Jk2 product band density of the gel scan image is quantified and the ratio between the densitometry value of the PCR product band detected from cells grown in the presence of EMF and the corresponding one without field exposure (EMF+/EMF-, Cell Recomb. Fr., Figs. 3, 4, 5) reports the changes in V(D)J recombination occurred upon each cell treatment (IMA/GSK). To normalize for DNA extraction levels we performed similar PCR amplifications from the same amount of template DNA using a pair of primers H1fw and H1rv to specifically detect the histone H1 gene.

**$\gamma$ H2AX foci analysis for irradiation induced DNA damage cellular response.** Cells were grown under similar conditions with those described above for recombination assays. Additionally, a DNA DSBs control cell lot either uninduced or one for each RAG stimulus (IMA or GSK) was exposed to a quick 20 min X ray cumulative dose exposure of 1 Gray (X-ray irradiation with a slow rate 50-milligray /min with a Mevatron Primus 2D, 6MV, SIEMENS instrument) prior to their harvest. The samples were irradiated at 100 cm distance from the source axis, the field size being of 30  $\times$  30 cm. The dosimetry was performed using a water phantom (1 cm water depth). Symmetry and homogeneity were checked, the dose proved to be homogenous throughout the sample in the used plates. For all treatments, twenty minutes after harvest, instead of extracting DNA, the cells from each individual culture type were separately spread onto clean designated slide sets using a Cytospin Centrifuge. The cells were then fixed with paraformaldehyde, permeabilized with Triton X and then doubly stained with: (a) Hoechst 33342 dye (THERMO SCIENTIFIC) (for their nuclei-DNA total staining in blue) and (b) immunofluorescently with primary unlabeled anti  $\gamma$ H2AX antibodies of mouse antigen specificity complemented with secondary Cy2 labeled anti primary source antibodies (rat anti mouse IgG Cy2 detection antibodies-green)(both from SIGMA ALDRICH); to identify in green the DSB repairing  $\gamma$ H2AX foci<sup>40</sup>. The slides were examined with a fluorescence microscope (OLYMPUS BX60) with adequate filter for the fluorophores, and images of the nuclei and  $\gamma$ H2AX foci recorded with a camera connected to the microscope. The images were analyzed using specific analysis software to quantify the number of foci per each cell treatment type, and morphologically to indicate their level of dispersion or nuclear positioning (see Fig. 6).

*Western blot analysis* for endogenous RAG time course induction in pre-B cells (Supplemental material Fig. 2S) following IMA/GSK treatment was performed as previously described in our work using anti RAG1 and anti RAG2 mouse monoclonal antibodies (gift from Dr. David G. Schatz, Yale University), and control sample purified murine core RAG1(384–1040) and coreRAG2 (1–387) fused to Maltose binding protein (MBP-40kD) which were transiently expressed in co-transfected HEK293T cells<sup>52</sup> (source ATCC CRL-3216).

*Absorption spectra measurements* were made using two identical broadband (0.8–16 GHz) horn antennas facing each-other and placed at 1 m distance. The measurement subjected sample was placed in close proximity (1 cm) of the receiver whereas the emission antenna (supplementary Fig. 4S a and b), was coupled to the generator. The receiver antenna was connected to a commercial Spectrum Analyzer (Keysight-AGILENT-HP N9935A, 0.1- 9 GHz) on which the received signals were recorded and analyzed. The shown absorption spectra in Fig. 2 were obtained after subtraction of the background spectra with no liquid sample placed in the container in front of the receiver antenna. The deionized water used for measurement has the conductivity < 5  $\mu$ S/cm, whereas the used unfiltered tap water with ions has the following characteristic measured chemical parameters per liter (l) pH 6.5–9.5, Conductivity < 800  $\mu$ S/cm, ammonia < 0.5 mg/l, free residual Chlorine < 0.5 mg/l, Fe < 200  $\mu$ g/l, Mn < 50  $\mu$ g/l, Al < 200  $\mu$ g/l, nitrites < 0.5 mg/l, nitrates < 50 mg/l, Borate salts 1 mg/l, Chlorides 250 mg/l, Sulphates 250 mg/l, 65 mg/l calcium carbonate, Hardness < 5degrees (dGH).

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## Author contributions

E.I. has performed all the cell irradiation experiments, maintained the pre-B cells in culture, performed PCR from genomic DNA and quantified the amplification results. A.M. and M.S. have performed all the experiments to measure the EMF parameters used in irradiation, absorption spectra, power flux density and electric field intensity measurements. M.T. has performed the experiments to detect and quantify the  $\gamma$ H2AX foci whereas D.S. helped in interpreting the results of their foci/cell analysis. M.C. has designed the experiments, performed the analysis and interpretation of the cell irradiation experiments, supervised experiments and wrote the manuscript. All authors reviewed the manuscript.

## Competing interests

The authors declare no competing interests.

## Additional information

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